Total Synthesis of the Cytotoxic Anhydrophytosphingosine Pachastrissamine (Jaspine B)

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Supporting Information

ABSTRACT: A short, 8-step synthesis of the marine natural product pachastrissamine has been developed that relies on a diastereoselective aldol reaction between a suitably protected hydantoin and an optically enriched α -chloroaldehyde. This synthetic route provides new opportunities for exploring structure activity relationships within this family of natural products.

P achastrissamine (1), also known as jaspine B, is a naturally occurring anhydrophytosphingosine that possesses a tetrahydrofuran core¹ and was first isolated from the Okinawan marine sponge *Pachastrissa* sp. in 2002 by Higa.² More recently, Debitus and co-workers reported the isolation of 1 along with a related anhydrophytosphingosine from the Vanuatuan marine sponge *Jaspis* sp. and assigned these compounds the common names jaspine A (2) and jaspine B (1) (Figure 1).³ The all-*cis*



Figure 1. Anhydrophytosphingosines 1 and 2 and furanodictine A (3).

trisubsituted tetrahydrofuran ring and absolute configuration (2S,3S,4S) of 1 were assigned by a combination of detailed NMR spectroscopic studies, mass spectral analysis, and chemical derivatization (e.g., formation of MTPA monoamides).² Importantly, pachastrissamine exhibited submicromolar cytotoxicity against P388, A549, HT29, MEL28, and HeLa cell lines and represents a potential lead for cancer therapy.²⁻⁴ More recent studies demonstrated that pachastrissamine inhibits the activity of sphingomyelin synthase and consequently increases intracellular ceramide levels, resulting in apoptosis in tumor cells by a caspase-dependent pathway.⁵ Owing to its potential importance as a lead for cancer therapy, considerable effort has been devoted to the synthesis of pachastrissamine.⁶⁻⁸ Presently, more than 25 syntheses of 1 have been reported; however, many of the synthetic strategies are lengthy (ranging from 9 to 19 steps) and have limited ability to access analogues or related natural products (e.g., furanodictine A(3)⁹ due to their reliance on carbohydrates or amino acids as chiral pool starting materials.

Previously, we have demonstrated that 1,2-*anti*-configured β ketochlorohydrins can be accessed in a straightforward manner



from the reaction of lithium enolates derived from methyl ketones with α -chloroaldehydes.¹⁰ In a single example, it was also shown that microwave-assisted heating the β -amidochlorohydrin **4** in a mixture of CH₃OH–H₂O afforded the β -hydroxy- γ -lactone **5** in excellent yield (Scheme 1).¹¹ While our

Scheme 1. Synthesis of γ -Lactone 5 and a Synthetic Strategy for Pachastrissamine (1) and Analogues



efforts to date have focused almost exclusively on the reaction of enolates derived from methyl ketones with α -chloroaldehydes,¹² we endeavored to extend this methodology to the preparation of aminotetrahydrofuranols, including pachastrissamine (1). As outlined in Scheme 1, it was envisaged that the reaction of a conformationally constrained α -aminoenolate (e.g., **6**) with an α -chloroaldehyde (e.g., 7) would give rise to an *anti-anti*-aminochlorohydrin (e.g., **8**) based on Evans-Cornforth type stereodirecting effect of the chlorine atom¹³ and the progression of the reaction through a chairlike 6-membered ring transition structure.¹⁴ A subsequent thermal cyclization would then provide direct access to α -amino- β -hydroxy- γ lactones (e.g., **9**) and serve as a launching point for the preparation of pachastrissamine and analogues of this potentially important natural product. Notably, and in contrast to chiral pool syntheses of pachastrissamine, this approach

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would rely on a single chlorine atom introduced via organocatalytic asymmetric α -chlorination¹² to control the relative stereochemistry of the amino alcohol function in **1**.¹⁵

As summarized in Table 1, these studies initiated with an investigation of the lithium aldol reaction between a small





^{*a*}Conditions: (A) LDA, THF, -78 °C, 30 min, then **19**; (B) LiHMDS, THF, -78 °C, 30 min, then **19**. ^{*b*}Yield and diastereomeric ratio determined by analysis of ¹H NMR spectra recorded on crude reaction products with internal standard. ^{*c*}The relative stereochemistry was not determined. ^{*d*}No product detected. ^{*e*}Products partially decompose during purification by flash column chromatography. ^{*f*}Major diastereomer is **23** (isolated in 45% yield).

collection of α -amino lactams/lactones **10-16**¹⁶ and α chloropentanal (**19**).^{12a} As indicated in entry 1, treatment of the TBS-protected hydantoin **10** with freshly prepared lithium diisopropylamide, followed by slow addition of the α chloroaldehyde **19** afforded the desired aldol adduct in 50% yield albeit with low diastereoselectivity (dr = 1:1). Unfortunately, coupling of the lithium enolates derived from compounds **11** or **12** with α -chloroaldehyde **19** under the same conditions failed to provide detectable amounts of the corresponding chlorohydrin (entries 2 and 3). Similarly, reaction of the protected diketopiperazines 13 and 14 with 19 provided only small amounts (<10%) of the corresponding aldol adducts (entries 4 and 5). Based on the modest success obtained with the TBS protected hydantoin 10, we next evaluated the benzyl- and Boc-protected analogues 15 and 16, respectively. As indicated in entries 6 and 7, these hydantoin derivatives reacted with α -chloropentanal to provide the desired β -amidochlorohydrins 22 and 23 in good yield and diastereoselectivity. In the latter example, formation of 23 also involves a 1,3-migration of a Boc protecting group¹⁷ and the β -amidochlorohydrin 23 could be isolated in good yield. It is noteworthy that this straightforward strategy for the synthesis of 23 could also be exploited for the preparation of various β hydroxy- α -amino acids by radical reduction of the chloromethine function,¹⁸ or more elaborate amino acid derivatives through displacement of the chloride with nitrogen or oxygen nucleophiles.

Following the successful preparation of the β -amidochlorohydrins **20**, **22**, and **23**, we next investigated the thermal cyclization of these compounds.¹¹ Fortunately, while attempts to effect cyclization of the β -amidochlorohydrins **20** or **22** at temperatures ranging from 50 °C to reflux in a variety of solvents (e.g., H₂O, CH₃OH, DMSO, MeCN, pH 7 buffered H₂O) led largely to their decomposition,¹¹ heating of the β amidochlorohydrin **23** in a mixture of CH₃OH–H₂O afforded the γ -lactone **27**, which could be isolated in 15% yield along with the fully deprotected β -amidochlorohydrin **28** (Table 2,

Table 2. Reaction Conditions for Formation of the γ -Lactone 27 from 23

entry	solvent(s)	$temp^a$ (°C)	27:28 ^{b} (% yield) ^{c}
1	H ₂ O-CH ₃ OH (1:1)	60	1:3 (62)
2	dimethyl carbonate	60	nd ^d
3	DMSO	60	nd ^d
4	H ₂ O-CH ₃ OH (1:1)	100	1.7:1
5	$H_2O^{-t}BuOH$ (1:1)	100	1.1:1
6	$H_2O-CF_3CH_2OH$ (1:1)	100	1.2:1
7^e	H ₂ O-CH ₃ OH (1:1.2)	100	2.7:1 (58)

"Reaction carried out in a sealed tube in an oil bath. ^bDetermined by analysis of ¹H NMR spectra recorded on crude reaction mixture. ^cCombined isolated yield. ^dNo product detected. ^eReaction carried out in a microwave reactor.

entry 1). Surprisingly, we were unable to convert the chloroamide 28 into the corresponding lactone even after heating at reflux for more than 24 h in H₂O, which resulted in complete recovery of the starting material. Together, these results suggest that formation of the desired γ -lactone 27 most likely proceeds via initial deprotection of the Boc-protected alcohol in 23 to provide the chlorohydrin 24 (Scheme 2). A second deprotection would then afford the chloroamide 28, while a structural rearrangement from the hydantoin 24 to carbamate 25 is a necessary step preceding the formation of γ lactone 27. In an effort to optimize the fortuitous rearrangement of hydantoin 24 to carbamate 25 necessary for the formation of lactone 27, we screened a variety of solvents and temperatures for this reaction (Table 2). As indicated in entries 2 and 3, heating of chlorohydrin 23 in dimethyl carbonate or DMSO at 60 °C afforded none of the desired γ -lactone 27. Interestingly, formation of the γ -lactone 27 was significantly enhanced when the reaction was heated in a mixture of Scheme 2. Potential Mechanism for the Formation of γ -Lactone 27



CH₃OH-H₂O at 100 °C (entry 4). Further increases to the reaction temperature (eg., 120 °C in a microwave reactor) in CH₃OH-H₂O (1:1) did not alter the ratio of γ -lactone 27 to β amidochlorohydrin 28. To further examine this process, a series of solvent mixtures were examined. As indicated in entries 5 and 6, heating the β -amidochlorohydrins 23 in H₂O-^tBuOH or H₂O-CF₃CH₂OH under the same reaction conditions failed to improve on the results obtained in CH₃OH-H₂O (1:1). After screening the reaction in several different mixtures of CH₃OH-H₂O, we eventually found that the ratio of γ -lactone 27 to β amidochlorohydrin 28 could be optimized to 2.7:1 in a H₂O-CH₃OH mixture of 1:1.2 (entry 7). This latter reaction could be carried out in a microwave reactor and was complete in 20 min, affording the desired γ -lactone 27 in 40% yield accompanied with a 18% yield of the deprotected β amidochlorohydrin 28 (entry 7).

Having developed a short synthetic route to the γ -lactone 27, we focused our efforts on applying this strategy to the total synthesis of (+)-pachastrissamine (1). As depicted in Scheme 3, the synthesis of 1 commenced with the asymmetric α chlorination of 6-heptenal (29) following the procedure reported by MacMillan.^{12c} While the chlorination of hexadecanal would eliminate two steps from the total synthesis (see below), the choice of 6-heptenal provides opportunities for the late stage production of pachastrissamine analogues through cross metathesis.¹⁹ A subsequent aldol reaction between the Boc-protected hydantoin 16 and the optically enriched α chloroaldehyde 30 afforded a mixture of chlorohydrins in good yield (68%) and diastereoselectivity (dr = 10:1:1:1), from which the desired aldol adduct could be isolated in 52% yield. Cyclization of the chlorohydrin 32 using the optimized reaction conditions (Table 2, entry 7) produce a 2.7:1 mixture of the desired the γ -lactone 33 and the chlorohydrin 34, in 60% yield. Cross-metathesis of the purified the γ -lactone 33 with 1undecene catalyzed by the second-generation Grubbs-Hoveyda catalyst²⁰ afforded the corresponding tetradecene, which was directly hydrogenated to give the lactone 35 in 43% yield over the two steps. Reduction of the lactone function involved treatment of compound 35 with DIBAL-H followed by reaction of the resulting mixture of lactols with Et₃SiH and $BF_3 \cdot OEt_2^{21}$ to afford the cyclic carbamate 37.^{6b} Finally, the carbamate was removed under basic conditions to afford (+)-pachastrissamine (1) in 91% yield. The spectroscopic data (¹H NMR, ¹³C NMR, HRMS, and IR) of our synthetic pachastrissamine (1) were in complete agreement with those reported in the literature,⁶ and the specific rotation was consistent with that reported ($[\alpha]_D = +10$ (c = 0.48 CHCl₃)).⁶ To further demonstrate the potential of this synthetic route for



providing access to structural analogues of pachastrissamine, the γ -lactone 33 was also converted into the truncated derivative 39 (i.e., dodecyl analogue) following an identical sequence of reactions that initiated with cross metathesis of 33 with 1-nonene (see inset, Scheme 3).

In conclusion, we have developed a short (eight step, 5% overall yield) asymmetric synthesis of pachastrissamine (1) that relies on the diastereoselective aldol reaction between a Bocprotected hydantoin and an optically enriched α -chloroaldehyde to secure the correctly configured core of 1. A fortuitous Boc protecting group migration proved key in the thermal cyclization of this material, which proved to be extremely sensitive to both temperature and solvent. Notably, the present synthesis compares well with those reported and, as demonstrated through the production of the dodecyl analogue 39, should be readily adapted to the production of analogues of pachastrissamine through cross-metathesis or initiation of the process with other readily available α -chloroaldehydes (e.g., 27, Scheme 2). Furthermore, the ready access to optically enriched β -amidochlorohydrins afforded by this process may also prove useful for the production of β -hydroxyamino acids and other more elaborate amino acid derivatives.

EXPERIMENTAL SECTION

Preparation of (\pm)-Hydantoin 23. To a cold (-78 °C), stirred solution of hexamethyldisilazane (1.2 mL, 5.5 mmol) in THF (4.0

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mL) was added n-butyllithium (2.59 M solution in hexanes, 2.0 mL, 5.1 mmol) dropwise. The resulting mixture was allowed to warm to rt slowly over 15 min and then added dropwise via cannula to a cold (-78 °C), stirred solution of Boc-protected hydantoin 16 (1.4 g, 4.6 mmol) in THF (34 mL). After the mixture was stirred for 45 min, (±)-2-chloropentanal (19) (674 mg, 5.5 mmol) in THF (4.0 mL) was added dropwise, and the resulting mixture was stirred for an additional 1.2 h. After this time, the reaction was treated with saturated aqueous solution of sodium dihydrogen phosphate (10 mL) and diluted with EtOAc (60 mL) and H₂O (60 mL). The phases were separated, and the aqueous phase was extracted with EtOAc (3 \times 50 mL). The combined organic phases were washed with brine (50 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude product by flash chromatography (silica gel, hexanes-EtOAc 3:1) afforded hydantoin 23 (876 mg, 45%) as a colorless oil. The stereochemical assignment for compound 23 was based on analysis of 1D NOESY spectra and comparison of its spectral data with that recorded on the structurally related hydantoin 35 (see below). ¹H NMR (400 MHz, $CDCl_{2}$) δ : 5.68 (br s, 1H), 5.11 (dd, 1H, I = 5.2, 2.1 Hz), 4.39 (dd, 1H, J = 2.1, 1.5 Hz), 4.14 (m, 1H), 1.81–1.64 (m, 2H), 1.57 (s, 9H), 1.53–1.41 (m, 2H), 0.94 (t, 3H, J = 7.8 Hz). ¹³C NMR (150 MHz, CDCl₃) *b*: 167.8, 152.2, 152.1, 145.6, 85.7, 84.1, 75.3, 61.2, 56.5, 36.0, 27.7, 27.5, 19.3, 13.3. IR (neat): 3312, 2980, 2936, 2876, 1813, 1774, 1725, 1274, 1153 cm⁻¹. HRMS: [M + Na]⁺ calcd for C₁₈H₂₉ClN₂NaO₇ 443.1556, found 443.1584.

Preparation of (\pm) - γ -Lactone 27. To an 80 mL vial containing hydantoin 23 (876 mg, 2.1 mmol) was added 1:1.2 mixture of deionized H₂O-CH₃OH (1:1.2, 25.6 mL), and the vial was sealed in a CEM Discover LabMate microwave reactor. The reaction mixture was then heated for 20 min at 100 °C (as monitored by a vertically focused IR temperature sensor). After this time, the mixture was diluted with EtOAc (30 mL) and H₂O (30 mL), and the phases were separated. The aqueous phase was extracted with EtOAc (3×30 mL), and the combined organic phases were washed with brine (30 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude product by flash chromatography (silica gel, CH₂Cl₂-CH₃OH 20:1) afforded γ -lactone 27 as a white solid (157 mg, 40%, mp = 136–138 °C) and hydantoin 28 (82 mg, 18%). Data for γ -lactone 27. ¹H NMR (500 MHz, CDCl₃) δ : 5.68 (br s, 1H), 5.21 (dd, 1H, J = 4.5, 7.6 Hz), 4.66 (m, 1H), 4.44 (d, 1H, J = 7.5 Hz), 1.96–1.81 (m, 2H), 1.57–1.49 (m, 2H), 1.01 (t, 3H, J = 7.4 Hz). ¹³ C NMR (150 MHz, CDCl₃) δ : 172.5, 156.7, 82.2, 77.0, 54.9, 30.9, 18.5, 13.7. IR (neat): 3268, 2962, 2877, 1753, 1710, 1222, 968 cm⁻¹. HRMS: $[M + H]^+$ calcd for $C_8H_{12}NO_4$ 186.0761, found 186.0741. Data for hydantoin 28. ¹H NMR (600 MHz, DMSO) δ : 5.87 (d, 1H, J = 7.5 Hz), 4.35 (t, 1H, J = 1.4 Hz), 4.00 (dt, 1H, J = 2.7, 9.8 Hz), 3.69 (ddd, 1H, J = 1.4, 7.5, 9.4 Hz), 1.54 (m, 2H), 1.36 (m, 2H), 0.90 (t, 3H, J = 7.2 Hz). ¹³ C NMR (150 MHz, DMSO) δ: 174.8, 158.1, 72.3, 62.2, 60.3, 35.3, 18.5, 13.3. IR (neat): 3263, 2920, 1768, 1716, 1261, 1147 cm⁻¹. HRMS: [M + H]⁺ Calcd for C₈H₁₄ClN₂O₃ 221.0725; Found 221.0719.

Preparation of (+)-(R)-2-Chlorohept-6-enal (30). To a cold (-10 °C), stirred solution of LiCl (1.91 g, 0.045 mol), copper(II) trifluoroacetate hydrate (4.36 g, 0.015 mol), sodium persulfate (7.18 g, 0.030 mol), and H_2O (1.2 mL, 0.066 mol) in acetonitrile (130 mL) was added imidazolidinone catalyst 31 (1.72 g, 6.0 mmol). After the mixture was stirred for 5 min, 6-heptenal (29) (3.38 g, 0.030 mol) in acetonitrile (20 mL) was added dropwise. The resulting mixture was stirred at -10 °C for 2 days, after which EtOAc (70 mL) and H₂O (70 mL) were added. The phases were separated, and the aqueous phase was washed with EtOAc (3 \times 70 mL). The combined organic phases were washed with brine (70 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude product by flash chromatography (silica gel, 1:1 CH₂Cl₂-pentane) afforded (+)-(R)-2-chlorohept-6-enal (30) as a colorless oil (2.60 g, 60%, ee >99.0%). $[\alpha]^{25}_{D}$: +37 (c 1.0, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ : 9.49 (d, 1H, J = 2.4 Hz), 5.77 (m, 1H), 5.03 (m, 1H), 5.00 (m, 1H), 4.16 (ddd, 1H, J = 8.1, 5.4, 2.4 Hz), 2.10 (m, 2H), 2.00 (m, 1H), 1.84 (m,1H), 1.64 (m, 1H), 1.55 (m, 1H). ¹³C NMR (150 MHz, CDCl₃) δ: 195.6, 137.9, 115.9, 64.2, 33.2, 31.7, 25.1. IR (neat): 2923, 2855, 1736, 1460, 913,

744 cm $^{-1}$. HRMS: $[M + H]^+$ calcd for $C_7H_{12}ClO$ 147.0566, found 147.0571.

Preparation of Hydantoin 32. To a cold (-78 °C), stirred solution of hexamethyldisilazane (1.4 mL, 6.6 mmol) in THF (6.0 mL) was added *n*-butyllithium (2.66 M solution in hexanes, 2.3 mL, 6.0 mmol) dropwise. The resulting mixture was allowed to warm to rt slowly over 15 min and then added dropwise via cannula to a cold (-78 °C), stirred solution of Boc-protected hydantoin 16 (1.65 g, 5.5 mmol) in THF (36 mL). After the mixture was stirred for 45 min, (+)-(R)-2-chlorohept-6-enal (30) (886 mg, 6.0 mmol) in THF (4.0 mL) was added dropwise, and the resultant mixture was stirred for an additional 1.2 h. After this time, the reaction mixture was treated with saturated aqueous sodium dihydrogen phosphate (10 mL) and diluted with EtOAc (60 mL) and H₂O (60 mL). The phases were separated, and the aqueous phase was extracted with EtOAc (3×50 mL). The combined organic phases were washed with brine (50 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude product by flash chromatography (silica gel, hexanes-EtOAc 3:1) afforded the hydantoin (32) as a white solid (1.28 g, 52%). $[\alpha]^{25}_{D}$: -12 (c 0.65, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ : 5.76 (m, 1H), 5.62 (br s, 1H), 5.12 (dd, 1H, J = 2.1, 5.2 Hz), 5.01 (m, 2H), 4.39 (br s, 1H), 4.12 (m, 1H), 2.10 (m, 2H), 1.84 (m, 1H), 1.74 (m, 1H), 1.57 (s, 9H), 1.51–1.54 (m, 2H), 1.46 (s, 9H). 13 C NMR (150 MHz, CDCl₃) δ 167.7, 152.2, 152.1, 145.6, 137.5, 115.6, 85.8, 84.1, 75.2, 61.3, 56.5, 33.4, 32.8, 27.8, 27.5, 25.1. IR (neat): 3307, 2981, 2936, 1813, 1774, 1725, 1370, 1274, 1153 cm⁻¹. HRMS: [M + Na]⁺ calcd for C₂₀H₃₁ClN₂NaO₇ 469.1712, found 469.1707.

Preparation of γ **-Lactone 33.** To the 80 mL vial containing hydantoin 32 (1.25 g, 2.7 mmol) was added a mixture of deionized H₂O-CH₃OH (1:1.2, 36 mL), and the vial was sealed in a CEM Discover LabMate microwave reactor. The reaction mixture was then heated for 20 min at 100 $^\circ \mathrm{C}$ (as monitored by a vertically focused IR temperature sensor). After this time, the mixture was diluted with EtOAc (60 mL) and H₂O (60 mL), and the phases were separated. The aqueous phase was extracted with EtOAc (3×50 mL), and the combined organic phases were washed with brine (50 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude product by flash chromatography (silica gel, CH₂Cl₂-CH₃OH 20:1) afforded the γ -lactone 33 as an off-white solid (250 mg, 44%; mp = 108–111 °C). $[\alpha]^{20}_{D}$: -13 (c 1.0, CHCl₃). ¹H NMR (600 MHz, CDCl₃) δ : 5.79 (m, 1H), 5.63 (s, 1H), 5.21 (dd, 1H, J = 4.5, 7.4 Hz), 5.02 (m, 2H), 4.65 (m, 1H), 4.44 (dd, 1H, J = 0.71, 7.4 Hz), 2.15 (m, 2H), 1.95 (m, 1H), 1.88 (m, 1H), 1.60 (m, 2H). ¹³ C NMR (150 MHz, CDCl₃) δ : 172.2, 156.3, 137.5, 115.6, 82.3, 76.9, 54.7, 33.2, 28.4, 24.4. IR (neat): 3251, 2925, 1774, 1726, 1365, 1218, 1110, 964, 655 cm⁻¹. HRMS: [M + Na]⁺ calcd for $C_{10}H_{13}NNaO_4$ 234.0737, found 234.0763.

Preparation of \gamma-Lactone 35. To a stirred solution of 1-undecene (0.24 mL, 1.2 mmol) in CH₂Cl₂ (7.5 mL) was added γ -lactone 33 (31 mg, 0.15 mmol) in CH₂Cl₂ (1.0 mL) followed by the second-generation Grubbs–Hoveyda catalyst (17 mg, 0.027 mmol). Nitrogen was bubbled through the mixture for 5 min, after which time the mixture was heated to reflux and maintained at this temperature for 3 h. The reaction mixture was then cooled to rt, filtered through a pad of Celite, and concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 1:1 hexanes–EtOAc) to give the cross-metathesis product as an off-white solid (25 mg). ¹H NMR (400 MHz, CDCl₃) δ : 6.28 (br s, 1H), 5.55–5.29 (m, 2H), 5.20 (dd, 1H, J = 4.2, 7.0 Hz), 4.65 (m, 1H), 4.47 (d, 1H, J = 7.3 Hz), 2.28–1.79 (m, 6H), 1.66–1.46 (m, 2H), 1.46–1.23 (m, 16H), 0.88 (t, 3H, J = 6.7 Hz).

To a stirred solution of the cross-metathesis product (25 mg, 0.073 mmol) in EtOAc–CH₃OH (1:1, 3.6 mL) was added Pd(OH)₂ (5.4 mg). Hydrogen was bubbled through the mixture for 30 min, and then the reaction was kept under hydrogen atmosphere for 1.5 h. After this time, the mixture was filtered through a pad of Celite and concentrated to afford γ -lactone **35** as an off-white solid (22 mg, 43% over two steps; mp = 146–148 °C). ¹H NMR (400 MHz, CDCl₃) δ : 5.97 (br s, 1H), 5.21 (dd, 1H, *J* = 4.4, 7.4 Hz), 4.65 (m, 1H), 4.45 (d, 1H, *J* = 7.4 Hz), 1.99–1.80 (m, 2H), 1.48 (m, 1H), 1.41–1.20 (m, 23H), 0.88 (t, 3H, *J* = 6.5 Hz). ¹³C NMR (100 MHz, CDCl₃) δ : 172.4, 156.6, 82.6,

77.0, 54.9, 31.9, 29.7, 29.7, 29.6, 29.6, 29.6, 29.6, 29.5, 29.3, 29.2, 29.0, 25.2, 22.7, 14.1. IR (neat): 3260, 2957, 2918, 2850, 1778, 1724, 1217 cm⁻¹. HRMS: $[M + H]^+$ calcd for $C_{19}H_{34}NO_4$ 340.2482, found 340.2462.

Preparation of Lactol 36. To a stirred solution of γ -lactone 35 (31 mg, 0.090 mmol) in CH₂Cl₂ (4.4 mL) at -55 °C was slowly added a solution of diisobutylaluminum hydride (1.0 M in THF, 0.32 mL, 0.32 mmol), and the resulting mixture was stirred for 12 h. An additional amount of diisobutylaluminum hydride (1.0 M in THF, 0.32 mL, 0.32 mmol) was then added, and the resulting mixture was stirred for 9 h at -55 °C. The excess reagent was then quenched by addition of 1 M HCl (1.5 mL), and the resulting mixture was diluted with EtOAc (10 mL), H₂O (5 mL), and brine (5 mL). The phases were separated, and the aqueous phase was extracted with EtOAc (3 \times 10 mL). The combined organic phases were washed with brine (10 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude material by flash chromatography (silica gel, EtOAc-hexanes 7:3) afforded lactol 36 as a white solid (26 mg, 85%, mp = 94–96 $^{\circ}$ C). ¹H NMR (400 MHz, CDCl₃) δ: 6.54 (br s, 1H), 5.30 (br s, 1H), 5.03 (dd, 1H, J = 3.6, 7.2 Hz), 4.27-4.20 (m, 2H), 3.46 (br s, 1H), 1.74 (m, 2H), 1.48–1.41 (m, 24H), 0.88 (t, 3H, J = 6.7 Hz). ¹³C NMR (100 MHz, CDCl₃) δ: 159.4, 100.9, 80.5, 80.1, 62.8, 31.9, 29.7, 29.6, 29.6, 29.6, 29.5, 29.3, 28.0, 26.0, 22.7, 14.1. IR (neat): 3428, 3304, 2918, 2850, 1762, 1733, 1071, 1014 cm⁻¹. HRMS: [M + H]⁺ calcd for C19H36NO4 342.2639, found 342.2650.

Preparation of Carbamate 37. To a cold (-78 °C), stirred solution of lactol 36 (26 mg, 0.076 mmol) and triethylsilane (0.12 mL, 0.76 mmol) in CH₂Cl₂ (3.6 mL) was added BF₃·OEt₂ (0.050 mL, 0.38 mmol), and the reaction mixture was allowed to warm to rt and stir for an additional 19.5 h. After this time, the reaction was treated with saturated aqueous solution of NaHCO3 (5 mL) and diluted with EtOAc (10 mL). The phases were separated, and the aqueous phase was washed with EtOAc $(3 \times 10 \text{ mL})$. The combined organic phases were washed with brine (10 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude material by flash chromatography (7:3 EtOAc-hexanes) provided the carbamate 37 as a white solid (8.0 mg, 32%; mp = 112-115 °C). Additional amounts of the starting lactol 36 (11 mg) were also isolated and redissolved in CH₂Cl₂ (5.0 mL). The resulting solution was cooled to 0 °C, and both Et₃SiH (0.10 mL, 0.63 mmol) and BF₃·OEt₂ (0.040 mL, 0.30 mmol) were added. The mixture was warmed to rt and allowed to stir for 48 h, after which time additional Et₃SiH (0.10 mL, 0.63 mmol) and BF₃. OEt₂ (0.040 mL, 0.30 mmol) were added. After a further 48 h, the reaction mixture was treated with saturated aqueous NaHCO₃ (5 mL) and diluted with EtOAc (10 mL). The phases were separated, and the aqueous phase was washed with EtOAc (3×10 mL). The combined organic phases were washed with brine (10 mL), dried (MgSO₄), filtered, and concentrated to give tetrahydrofuran carbamate 37 as a white solid (9.5 mg, 38%). Total isolated yield of carbamate 37 = 70%. $[\alpha]^{20}_{D}$: +54 (c 0.72, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ : 6.06 (br s, 1H), 4.95 (dd, 1H, J = 3.6, 7.4 Hz), 4.37 (dd, 1H, J = 3.6, 7.4 Hz), 3.94 (d, 1H, J = 10.4 Hz), 3.54–3.48 (m, 2H), 1.83–1.71 (m, 2H), 1.51-1.20 (m, 24H), 0.88 (t, 3H, J = 6.6 Hz). ¹³C NMR (150 MHz, CDCl₃) *b*: 159.2, 83.2, 80.9, 73.3, 57.1, 31.9, 29.7, 29.7, 29.7, 29.6, 29.6, 29.6, 29.6, 29.5, 29.3, 28.1, 26.0, 22.7, 14.1. IR (neat): 3330, 3242, 2921, 2848, 1756, 1720, 1070 cm⁻¹. HRMS: [M + H]⁺ calcd for C19H36NO3 326.2690, found 326.2686.

Preparation of (+)-Pachastrissamine (1). To a stirred solution of tetrahydrofuran carbamate 37 (7.1 mg, 0.022 mmol) in EtOH (1.5 mL) was added a solution of KOH (1 M in H₂O, 1.2 mL), and the mixture was heated to reflux (85 °C) and maintained at this temperature for 15 h. The reaction mixture was then cooled to rt and diluted with EtOAc (10 mL) and H₂O (5 mL). The phases were separated, and the aqueous layer was extracted with EtOAc (4 × 10 mL). The combined organic layers were washed with brine, dried (MgSO₄), and concentrated. Purification of crude product by flash chromatography (silica gel, CHCl₃–CH₃OH–NH₄OH 95:8:1) afforded (+)-pachastrissamine (1) as a white solid (5.9 mg, 91%; mp = 83–85 °C, lit.^{6c} mp = 89–91 °C). $[\alpha]^{20}_{Di}$: +10 (*c* 0.48, CHCl₃). $[\alpha]^{20}_{Di}$: +16 (*c* 0.45, EtOH). ¹H NMR (600 MHz, CDCl₃) δ : 3.92 (dd,

1H, J = 7.7, 7.7 Hz), 3.87 (m, 1H), 3.73 (m, 1H), 3.65 (m, 1H), 3.52 (dd, 1H, J = 7.7, 7.7 Hz), 2.25 (br s, 2H), 1.72–1.59 (m, 2H), 1.47–1.19 (m, 24H), 0.88 (t, 3H, J = 6.8 Hz). ¹³C NMR (150 MHz, CDCl₃) δ : 83.2, 72.3, 71.7, 54.3, 31.9, 29.8, 29.7, 29.7, 29.6, 29.6, 29.6, 29.4, 29.4, 29.3, 26.3, 22.7, 14.1 cm⁻¹. IR (neat): 3341, 2917, 2849, 1583, 1470, 1034 cm⁻¹. HRMS: [M + H]⁺ calcd for C₁₈H₃₈NO₂ 300.2897, found 300.2900.

Preparation of (\pm) - γ -Lactone 38. To a stirred solution of γ lactone 33 (20 mg, 0.094 mmol) in CH₂Cl₂ (5.0 mL) was added 1nonene (0.13 mL, 0.76 mmol) followed by the second-generation Grubbs-Hoveyda catalyst (12 mg, 0.019 mmol). The reaction mixture was heated to reflux and maintained at this temperature for 3 h, after which time the mixture was filtered through a pad of Celite and the filtrate was concentrated under reduced pressure. The residue was purified by flash chromatography (silica gel, 1:1 hexanes-EtOAc) to give the γ -lactone 38 as a white solid (17 mg, 54%, mp = 158–161 °C). ¹H NMR (600 MHz, CDCl₃) δ : 5.59 (s, 1H), 5.46–5.34 (m, 2H), 5.20 (dd, 1H, J = 4.4, 7.3 Hz), 4.65 (m, 1H), 4.44 (d, 1H, J = 7.3 Hz), 2.10-1.84 (m, 7H), 1.61-1.51 (m, 3H), 1.35-1.23 (m, 14H), 0.87 (t, 3H, J = 6.8 Hz). ¹³C NMR (150 MHz, CDCl₃) δ : 172.2, 156.3, 131.9, 128.6, 82.4, 77.0, 54.9, 32.5, 32.0, 31.8, 29.5, 29.1, 29.1, 28.4, 25.1, 22.6, 14.1. IR (neat): 3344, 2959, 2923, 2854, 1784, 1751, 1242, 1103, 965 $\rm cm^{-1}$.

To a stirred solution of cross-metathesis product (17 mg, 0.050 mmol) in EtOAc–CH₃OH 1:1 (3.0 mL), was added Pd(OH)₂ (3.4 mg), and the resulting mixture was stirred under a hydrogen atmosphere (balloon) for 1.5 h. After this time, the mixture was filtered through a pad of Celite and concentrated. Purification of crude product by flash chromatography (silica gel, 1:1 hexanes–EtOAc) afforded the γ -lactone **38** (14 mg, 82%). ¹H NMR (600 MHz, CDCl₃) δ : 5.55 (b, 1H), 5.21 (dd, 1H, *J* = 4.5, 7.5 Hz), 4.64 (m, 1H), 4.44 (dd, 1H, *J* = 0.66, 7.5 Hz), 1.93 (m, 1H), 1.86 (m, 1H), 1.51–1.42 (m, SH), 1.40–1.34 (m, 3H), 1.32–1.23 (m, 16H), 0.88 (t, 3H, *J* = 6.8 Hz). ¹³C NMR (150 MHz, CDCl₃) δ : 172.2, 156.3, 82.5, 76.9, 54.8, 31.9, 29.6, 29.6, 29.5, 29.4, 29.3, 29.2, 28.9, 25.1, 22.7, 14.1. IR (neat): 3673, 3250, 2981, 2917, 2854, 1777, 1726, 1393, 1219, 1066 cm⁻¹. HRMS: [M + H]⁺ calcd for C₁₇H₃₀NO₄ 312.2200, found 312.2169.

Preparation of the (+)-Dodecyl Analogue of Pachastriss**amine (39).** To a cold (-78 °C), stirred solution of γ -lactone 38 (28 mg, 0.082 mmol) in CH₂Cl₂ (4.0 mL) was slowly added a solution of diisobutylaluminum hydride (1.0 M in THF, 0.29 mL, 0.29 mmol), and the resulting mixture was stirred for 7 h. The excess reagent was then guenched by addition of 1 M HCl (1.0 mL), and the resulting mixture was diluted with EtOAc (10 mL) and H₂O (10 mL). The phases were separated, and the aqueous phase was extracted with EtOAc (3×10 mL). The combined organic phases were washed with brine (10 mL), dried (MgSO₄), filtered, and concentrated. Purification of the crude material by flash chromatography (silica gel, EtOAchexanes 2:1) afforded the corresponding lactol (20.5 mg, 72%). $^1\mathrm{H}$ NMR (600 MHz, CDCl₃) δ: 5.66 (b, 1H), 5.31 (b, 1H), 5.03 (dd, 1H, J = 3.7, 7.2 Hz), 4.25 (m, 1H), 4.22 (d, 1H, J = 7.2 Hz), 2.64 (d, 1H, J = 7.9 Hz), 1.75 (m, 2H), 1.42 (m, 2H), 1.28 (m, 22H), 0.88 (t, 3H, J = 6.8 Hz). ¹³C NMR (150 MHz, CDCl₃) δ : 158.6, 101.0, 80.4, 80.2, 62.5, 31.9, 29.7, 29.6, 29.5, 29.5, 29.4, 29.3, 28.0, 25.9, 22.7, 14.1. IR (neat): 3341, 2921, 2852, 1736, 1075 cm⁻¹. HRMS: [M + H]⁺ calcd for C₁₇H₃₂NO₄ 314.2300, found 314.2319.

To a cold (-78 °C), stirred solution of lactol (22 mg, 0.064 mmol) and triethylsilane (0.10 mL, 0.64 mmol) in CH₂Cl₂ (3.0 mL) was slowly added BF₃·OEt₂ (40 μ L, 0.32 mmol), and the reaction mixture was allowed to warm to rt and stir for an additional 16 h. After this time, the reaction mixture was treated with saturated aqueous NaHCO₃ (5 mL) and diluted with EtOAc (10 mL). The phases were separated, and the aqueous phase was washed with EtOAc (3 × 10 mL). The combined organic phases were washed with brine (10 mL), dried (MgSO₄), filtered, and concentrated to provide the tetrahydrofuran carbamate (20 mg, 90%). ¹H NMR (500 MHz, CDCl₃) δ : 5.10 (s, 1H), 4.96 (dd, 1H, *J* = 3.7, 7.5 Hz), 4.37 (dd, 1H, *J* = 3.9, 7.5 Hz), 3.93 (d, 1H, *J* = 10.3 Hz), 3.55–3.50 (m, 2H), 1.77 (m, 2H), 1.47–1.23 (m, 24H), 0.88 (t, 3H, *J* = 6.8 Hz). ¹³C NMR (150 MHz, CDCl₃) δ : 158.5, 83.3, 80.9, 73.4, 56.9, 31.9, 29.7, 29.6, 29.6,

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29.5, 29.4, 29.3, 28.1, 26.0, 22.7, 14.1. IR (neat): 2923, 2854, 1747, 1718, 1072 cm⁻¹. HRMS: $[M + H]^+$ calcd for $C_{17}H_{32}NO_3$ 298.2400, found 298.2355.

To a stirred solution of tetrahydrofuran carbamate (12 mg, 0.037 mmol) in EtOH (1.7 mL) was added a solution of KOH (1 M in H₂O, 1.7 mL), and the mixture was heated to reflux and maintained at this temperature for an additional 12 h. The reaction mixture was then cooled to rt and concentrated. Purification of the crude product by flash chromatography (silica gel, CHCl₃–CH₃OH–NH₄OH 95:6:1) afforded (\pm)-dodecyl analogue of pachastrissamine (**39**) as a white solid (9.0 mg, 82%; mp = 74–76 °C). ¹H NMR (400 MHz, CDCl₃) δ : 3.92 (dd, 1H, *J* = 7.3, 8.4 Hz), 3.86 (dd, 1H, *J* = 3.5, 5.0 Hz), 3.73 (m, 1H), 3.66 (m, 1H), 3.51 (dd, 1H, *J* = 6.6, 8.4 Hz), 2.02 (br s, 2H), 1.69–1.61 (m, 2H), 1.42–1.22 (m, 24H), 0.88 (t, 3H, *J* = 6.8 Hz). ¹³C NMR (150 MHz, CDCl₃) δ : 83.2, 72.4, 71.7, 54.2, 31.9, 29.8, 29.7, 29.6, 29.6, 29.5, 29.5, 29.4, 29.3, 26.3, 22.7, 14.1. IR (neat): 3675, 3157, 2918, 2850, 1469, 1054 cm⁻¹. HRMS: [M + H]⁺ calcd for C₁₆H₃₄NO₂ 272.2600, found 272.2575.

ASSOCIATED CONTENT

Supporting Information

General experimental methods and copies of ¹H and ¹³C NMR spectra. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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